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07/18/97

LIMBACH & LIMBACH L.L.P.
2001 Ferry Building, San Francisco, CA 94111
(415) 433-4150

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Docket No. SNUS - 125
Anticipated Classification of
this Application:
Class: _____
Subclass: _____
Prior Application:
Examiner: G. Hollinden
Art Unit: 1211

CONTINUATION OR DIVISIONAL APPLICATION UNDER 37 CFR 1.60

Assistant Commissioner
for Patents
Box Patent Application
Washington, D.C. 20231

Sir:

CERTIFICATION UNDER 37 CFR 1.10

I hereby certify that this New Application and the documents referred to as enclosed herein are being deposited with the United States Postal Service on this date **July 18, 1997**, in an envelope bearing "Express Mail Post Office To Addressee" Mailing Label Number **EM393841454US** addressed to: Box Patent Application, Assistant Commissioner for Patents, Washington, D.C. 20231.

HOWARD WONG
(Name of person mailing paper) (Signature)

This is a request for filing a [X] continuation application under 37 CFR 1.60, of pending prior application no. 08/770,522 filed on December 20, 1996 of Steven C. Quay for METHOD OF ULTRASOUND IMAGING.

1. [X] Enclosed is a **COMPLETE COPY** of the prior application, including the oath or declaration as originally filed. A declaration verifying it as a true copy appears in ¶23 below. (See ¶13 for drawing requirements.)
2. Name of applicant(s) (as originally filed and as last amended) and current correspondence address of applicant(s):
Steven C. Quay,

3. [X] This application discloses and claims only subject matter disclosed in the prior application whose particulars are set out above and the inventor(s) in this application are
[X] the same
[] less than those named in the prior application and it is requested that the following inventor(s) identified above for the prior application be deleted:

4. [X] The inventorship for all the claims in this application are
[X] the same

☐ not the same, and an explanation, including the ownership of the various claims at the time the last claimed invention was made, is submitted.

5. ☒ A verified statement to establish small entity status under 37 CFR 1.9 and 1.27 ☐ is enclosed ☒ was filed in the prior application no. 07/936,011 filed on September 2, 1992 and such status is still proper and desired (37 CFR 1.28(a)).

6. ☒ The filing fee is calculated below:

CLAIMS AS FILED IN THE PRIOR APPLICATION LESS ANY CLAIMS
CANCELLED BY AMENDMENT BELOW

CLAIMS AS FILED						
	CLAIMS REMAINING AFTER AMENDMENT OF ¶10	CLAIMS ADDED BY PRELIMINARY AMENDMENT OF ¶11	TOTAL CLAIMS FILED	NUMBER EXTRA*	RATE	BASIC FEE \$770
Total Claims	0	20	20	-20 = 0	× 22 =	\$0.00
Independent Claims	0	4	4	-3 = 1	× 80 =	\$80.00
___ FIRST PRESENTATION OF MULTIPLE DEP CLAIM					+ 260 =	\$
TOTAL						\$850.00
Small Entity 50% Filing Fee Reduction (if applicable)						\$425.00

* If the difference is less than zero, enter "0."

7. ☒ A check in the amount of \$425.00 is enclosed.

8. **AUTHORIZATION TO CHARGE ADDITIONAL FEES**

☒ The Commissioner is hereby authorized to charge the following **ADDITIONAL** fees which may be required by this paper and during the entire pendency of this application to Account No. 12-1420. The Commissioner is hereby authorized to charge payment of any fees associated with this communication or credit any overpayment to Deposit Account No. 12-1420. **A duplicate copy of this sheet is enclosed.**

☒ 37 CFR 1.16 (filing fees)

☒ 37 CFR 1.16 (presentation of extra claims)

☐ 37 CFR 1.16(e) (surcharge for filing the basic filing fee and/or declaration on a date later than the filing date of the application)

☐ 37 CFR 1.17 (application processing fees)

☐ 37 CFR 1.18 (issue fee at or before mailing of Notice of Allowance, pursuant to 37 CFR 1.311(b)).

9. **INSTRUCTIONS AS TO OVERPAYMENT**

☒ credit Account No. 12-1420

☐ refund

10. ☐ Cancel in this application original Claims of the prior application before calculating the filing fee. (At least one original independent claim must be retained for filing purposes).

11. ☒ A preliminary amendment is enclosed. (Claims added by this amendment have been properly numbered consecutively beginning with the number next following the highest numbered originally claimed in the prior application.)

12. ☒ RELATE BACK - 35 USC 120: Amend the specification by inserting before the first line the sentence:

--This is a ☒ continuation of Application no. 08/770,522, filed December 20, 1996, which is a

continuation application of application No. 08/649,910, filed May 16, 1996 which is a continuation application of 08/380,085 filed January 30, 1995, now U.S. Patent No. 5,558,854 which is a divisional application of 07/936,011 filed September 2, 1992 which is a continuation-in-part application of 07/893,657 filed June 5, 1992, now U.S. Patent No. 5,409,688 which is a continuation-in-part application of 07/761,311 filed September 17, 1991.--

[Note to form user: lines for item 12 are intentionally spaced to permit Examiner amendments.]

13. DRAWINGS

- ☐ Transfer the drawings from the prior application to this application and abandon said prior application as of the filing date accorded this application. A duplicate copy of this sheet is enclosed for filing in the prior application file. (May only be used if signed by person authorized by 37 CFR 1.138 and before payment of base issue fee.)
- ☐ New formal drawings are enclosed.

14. PRIORITY

- ☐ Priority of application no. _ filed on _ in _ is claimed under 35 USC 119.
- ☐ The certified copy of the priority application has been filed in prior application no. _ filed on _.

15. ASSIGNMENT

- ☒ The prior application is assigned of record to Sonus Pharmaceuticals, Inc., 20336 20th Avenue S.E., Suite 102, Bothell, WA 98021;
Assignment recorded in PTO on September 2, 1993, Reel 6676 Frame(s) 0699.
- ☐ The prior application is assigned, and the assignment (copy attached) was submitted to PTO for recording on _.
- ☐ An assignment of the invention to is attached. A copy of Form PTO-1595 (Recordation Cover Sheet) is also attached.

16. ☒ The power of attorney in the prior application is to the members of the firm of LIMBACH & LIMBACH L.L.P., 2001 Ferry Building, San Francisco, California, 94111, a firm composed of:

Karl A. Limbach	18,689	Stephen M. Everett	30,050	Alan A. Limbach	39,749
George C. Limbach	19,305	Alfred A. Equitz	30,922	Douglas C. Limbach	35,249
John K. Uilkema	20,282	W. Patrick Bengtsson	32,456	Brian J. Keating	39,520
J. William Wigert, Jr.	24,582	Mark A. Dalla Valle	34,147	Seong-Kun Oh*	
Philip M. Shaw, Jr.	25,376	Charles P. Sammut	28,901	Mayumi Maeda	40,075
Neil A. Smith	25,441	Richard A. Nebb	33,540	Kent J. Tobin	39,496
Veronica C. Devitt	29,375	Richard E. Wawrzyniak	36,048	Michael R. Ward	38,651
Ronald L. Yin	27,607	Alan D. Minsk	35,956	Steven M. Santisi	40,157
Gerald T. Sekimura	30,103	Mark C. Pickering	36,239	J. Thomas McCarthy	22,420
Michael A. Stallman	29,444	Kathleen A. Frost	37,326	Philip D. Reilly	P-41,415
Philip A. Girard	28,848	Alan S. Hodes	38,185		
Michael J. Pollock	29,098	Patricia Coleman James	37,155		

* Recognition under 37 CFR 10.9(b)

- a. ☐ The power appears in the original papers in the prior application.
- b. ☐ Since the power does not appear in the original papers, a copy of the power in the prior application is enclosed.
- c. ☐ A new power has been executed and is attached.
- d. ☒ Address all future communications to LIMBACH & LIMBACH L.L.P., Attn: W. Patrick Bengtsson, 2001 Ferry Building, San Francisco, California, 94111.

17. STATEMENT UNDER 37 CFR 3.73(B) (certification of title in assignee, if applicable, see MPEP 324)
☐ A statement satisfying the requirements of 37 CFR 3.73(b)
☐ is attached.
☐ was filed in the prior application.
☐ A copy of the statement previously filed in the prior application is attached.
18. ☒ An Information Disclosure Statement is enclosed with Form PTO-1449 (modified) and ten references.
19. ☐ Enclosed is a Statement Requesting Deletion of Names of Persons Who are No Longer Inventors.
20. MAINTENANCE OF COPENDENCY OF PRIOR APPLICATION
(This item must be completed and the necessary papers filed in the prior application if the period set in the prior application has run).
☐ A petition, fee and response has been filed to extend the term in the pending prior application until _.
☐ A copy of the petition for extension of time in the prior application is attached.
21. CONDITIONAL PETITIONS FOR EXTENSION OF TIME IN PRIOR APPLICATION
(Complete this item and file conditional petition in prior application if previous item (20) not applicable).
☐ A conditional petition for extension of time is being filed in the pending prior application.
☐ A copy of the conditional petition for extension of time in the prior application is attached.
22. ABANDONMENT OF PRIOR APPLICATION
☐ Please abandon the prior application at a time while the prior application is pending or when the petition for extension of time or to revive in that application is granted and when this application is granted a filing date so as to make this application copending with said prior application.
23. ☒ I hereby verify that the attached papers are a true copy of prior complete application no. and no amendments referred to in the oath or declaration filed to complete the prior application introduced new matter therein.

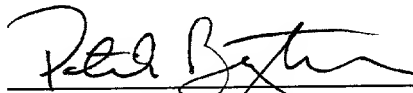
The undersigned declares further that all statements made herein of his or her own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

LIMBACH & LIMBACH L.L.P.

July 17, 1997
(Date)

Atty Docket: SNUS-125

By:


W. Patrick Bengtsson
Registration No. 32456
Attorney(s) or Agent(s) of Record

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of)	Group Art Unit:
STEVEN C. QUAY)	Examiner:
Appl. No. NEW)	<u>PRELIMINARY AMENDMENT</u>
Filed: HERewith)	2001 Ferry Building
For: METHOD OF ULTRASOUND))	San Francisco, CA 94111
IMAGING (As Amended)))	(415) 433-4150
)	

Assistant Commissioner
for Patents
Washington, D.C. 20231

Sir:

In advance of prosecution in the above-referenced application, please enter the following Amendments and remarks.

A M E N D M E N T S

In the Specification:

At page 1, please delete the present title and replace it with --Method of Ultrasound Imaging--.

In the claims:

Please cancel claims 1-14 and add new claims 15-34.

-- 15. In a method comprising ultrasound imaging, the improvement comprising enhancing the contrast in an ultrasound image by selecting for use as an enhancing agent microbubbles of a gas including perfluorohexane.

16. The method of claim 15 wherein said agent further comprises an aqueous solution of human protein.

17. The method of claim 15 wherein said agent further comprises an aqueous solution of liposomes.

18. The method of claim 15 wherein said agent further comprises an aqueous solution of microspheres.

19. The method of claim 15 wherein said agent further comprises a powder and diluent.

20. In a method comprising ultrasound imaging, the improvement comprising enhancing the contrast in an ultrasound image by selecting for use as an enhancing agent microbubbles of a gas including perfluoropentane.

21. The method of claim 20 wherein said agent further comprises an aqueous solution of human protein.

22. The method of claim 20 wherein said agent further comprises an aqueous solution of liposomes.

23. The method of claim 20 wherein said agent further comprises an aqueous solution of microspheres.

24. The method of claim 20 wherein said agent further comprises a powder and diluent.

25. In a method comprising ultrasound imaging, the improvement comprising enhancing the contrast in an ultrasound image by selecting for use as an enhancing agent microbubbles of a gas including perfluorobutane.

26. The method of claim 25 wherein said agent further comprises an aqueous solution of human protein.

27. The method of claim 25 wherein said agent further comprises an aqueous solution of liposomes.

28. The method of claim 25 wherein said agent further comprises an aqueous solution of microspheres.

29. The method of claim 25 wherein said agent further comprises a powder and diluent.

30. In a method comprising ultrasound imaging, the improvement comprising enhancing the contrast in an ultrasound image by selecting for use as an enhancing agent microbubbles of a gas including perfluoropropane.

31. The method of claim 30 wherein said agent further comprises an aqueous solution of human protein.

32. The method of claim 31 wherein said agent further comprises an aqueous solution of liposomes.

33. The method of claim 32 wherein said agent further comprises an aqueous solution of microspheres.

34. The method of claim 33 wherein said agent further comprises a powder and diluent. --

R E M A R K S

Entry of this supplemental preliminary amendment is respectfully requested. After entry of this amendment, claims 15-34 will be pending. An Information Disclosure Statement is also enclosed.

The present invention relates to the identification of certain gases for use in ultrasound contrast which have improved persistence in the body. The gases, of which fluorine-containing gases are preferred, can be used per se (as "free gas microbubbles") or can be adopted into known carriers such as albumin microspheres, solid particles or phospholipid liposomes.

More specific support for the new claims is as follows:

Claims 15, 20, 25 and 30 are supported at Table II, pages 28-29 which lists perfluoropropane,

(octafluoropropane), perfluorobutane (decafluorobutane) and perfluoropentane (dodecafluoropentane). Perfluorohexane is supported by adding one $-CF_2-$ to perfluoro-n-pentane as disclosed at page 27, lines 7-15 and in Example 6 at page 40.

Claims 16, 21, 26 and 31 are supported at page 17, lines 16-26.

Claims 17, 22, 27 and 32 are supported at page 15, lines 34-35 and page 16, lines 1-13.

Claims 18, 23, 28 and 33 are supported at page 12, lines 11-23.

Claims 19, 24, 29 and 34 are supported at page 15, lines 1-6.

Claims relating to these gases have already issued to the assignee of the present application as follows: U.S. Patent Nos. 5,393,524; 5,573,751; 5,409,688; 5,558,094 and 5,558,854.

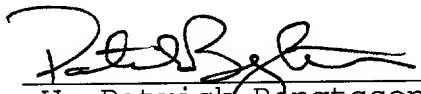
Applicant has the following cases pending in which there are claims which reference these gases as follows: 08/745,256; 08/710,849; 08/466,104; 08/646,910 and 08/770,522.

Applicant is prepared to provide the appropriate terminal disclaimers upon indication of allowable subject matter.

Early and favorable action is requested.

Respectfully submitted,

LIMBACH & LIMBACH L.L.P.

Dated: July 17, 1987 By: 
W. Patrick Bengtsson
Registration No. 32,456

Atty Docket: SNUS 125

Attorney for Applicant

PATENT

-1-

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of) Group Art Unit:
)
STEVEN C. QUAY) Examiner:
)
Serial No. NEW) PRELIMINARY AMENDMENT
)
Filed: December 20, 1996)
) 2001 Ferry Building
For: GASEOUS ULTRASOUND CONTRAST) San Francisco, CA 94111
MEDIA AND METHOD FOR) (415) 433-4150
SELECTING GASES FOR USE AS)
ULTRASOUND CONTRAST MEDIA) Docket No. SNUS 124

EXPRESS-MAIL CERTIFICATE

I hereby certify that this correspondence is being deposited with the United States Postal Service "Express Mail Service" Label No. 1B377890480US, postage paid in an envelope, addressed to: Commissioner of Patents and Trademarks, Washington, DC 20231 on 12/20/96

Dated: 12/20/96 By: Orig. signed
Name: By H. Wang

Honorable Commissioner of Patents
and Trademarks
Washington, D.C. 20231

Sir:

Prior to action in the above-referenced application, please enter the following amendment and remarks:

IN THE SPECIFICATION

At page 1, please delete the present title and replace it with --Contrast Media for Ultrasound Imaging--.

IN THE CLAIMS

Please cancel claims 1-14 and add new claims 15-50 as follows:

-- 15. A biocompatible ultrasound contrast media in powder form, the improvement comprising including a gaseous fluorine-containing chemical with said media.

16. The contrast agent of claim 15 wherein said fluorine-containing chemical is selected from the

group consisting of gaseous perfluoropropane, perfluorobutane, perfluoropentane, perfluorohexane, sulfur hexafluoride and mixtures thereof.

17. The contrast agent of claim 15 wherein said fluorine-containing chemical is gaseous perfluorohexane.

18. The contrast agent of claim 15 wherein said fluorine-containing chemical is sulfur hexafluoride.

19. The contrast agent of claim 15 wherein said gaseous fluorine-containing chemical is present in the form of microbubbles.

20. The contrast agent of claim 19 wherein a portion of said microbubbles are less than 8 microns in diameter.

21. The contrast agent of claim 19 wherein said microbubbles are formed by manual suspension.

22. A biocompatible ultrasound contrast agent comprising a sterile aqueous suspension of a gas emulsion of microbubbles of gaseous perfluoropropane, perfluorobutane, perfluoropentane, perfluorohexane, sulfur hexafluoride and mixtures thereof.

23. The contrast agent of claim 22 wherein microbubbles comprise perfluorohexane.

24. The contrast agent of claim 22 wherein said microbubbles comprise perfluoropropane.

25. The contrast agent of claim 22 wherein said microbubbles comprise sulfur hexafluoride.

26. The contrast agent of claim 22 wherein the aqueous suspension further comprises sodium chloride.

27. The contrast agent of claim 26 wherein said sodium chloride is present at a concentration of approximately 0.9 g per 100.0 mL.

28. The contrast agent of claim 22 further comprising amphipathic materials.

29. The contrast agent of claim 28, wherein said amphipathic materials include phospholipids.

30. The contrast agent of claim 22 wherein at least a portion of said microbubbles have a median diameter of between 2 and 5 microns.

31. The contrast agent of claim 22 wherein said microbubbles are present in said aqueous suspension at a density of greater than 50 million microbubbles per mL.

32. The contrast agent of claim 22 wherein said gas emulsion suspension comprises between 2 and 10% gas by volume.

33. A biocompatible ultrasound contrast agent containing a sterile gas-liquid system, wherein said gas comprises at least one gaseous fluorocarbon selected from the group consisting of the homologous series of non-branched fluorocarbons obtained from perfluoro-n-pentane by increasing or reducing the

number of -CF₂- groups present in the molecule and said liquid comprises water.

34. The contrast agent of claim 33 wherein said non-branched fluorocarbons are selected from the group consisting of gaseous perfluoropropane, perfluorobutane, perfluoro-n-pentane, perfluoro-n-hexane, and mixtures thereof.

35. The contrast agent of claim 33 wherein said gas is perfluoropropane.

36. The contrast agent of claim 33 wherein said gas is perfluorohexane.

37. The contrast agent of claim 33 wherein said gas is perfluoropentane.

38. The contrast agent of claim 33 wherein said gas-liquid system further comprises microspheres.

39. The contrast agent of claim 38 wherein said microspheres are formed from human protein.

40. The contrast agent of claim 33 wherein said gas-liquid system comprises an emulsion.

41. The contrast agent of claim 33 wherein said gas-liquid system comprises microscopic, spherical vesicles.

42. The contrast agent of claim 41 wherein said vesicles comprise amphipathic materials.

43. The contrast agent of claim 42 wherein said amphipathic materials include phospholipid.

44. The contrast agent of claim 33 wherein said gas-liquid system further comprises solid particles.

45. The contrast agent of claim 44 wherein at least a portion of said particles range in size from 5 to 10 microns.

46. Biocompatible ultrasound contrast agent comprising a sterilized mixture including microspheres and a gaseous fluorine-containing chemical.

47. The contrast agent of claim 46 wherein said fluorine-containing chemical is selected from the group consisting of perfluoropropane, perfluorobutane, perfluoro-n-pentane, perfluoro-n-hexane, sulfur hexafluoride and mixtures thereof.

48. The contrast agent of claim 47 wherein said fluorine-containing chemical comprises perfluoropropane.

49. The contrast agent of claim 48 wherein said microspheres contain human protein.

50. A biocompatible ultrasound contrast agent including nitrogen microbubbles, the improvement comprising including a fluorinated gas in said microbubbles.

51. The contrast agent of claim 50 wherein said fluorinated gas is selected from the group

consisting of perfluoropropane, perfluorobutane, perfluoro-n-pentane, perfluoro-n-hexane, sulfur hexafluoride and mixtures thereof.

52. The contrast agent of claim 50 wherein at least a portion of said microbubbles are less than 8 microns in size. --

REMARKS

Entry of this amendment is respectfully requested. Claims 1-14 are canceled. New claims 15-52 are added.

These new claims are supported in applicant's specification as follows:

The present invention relates to the identification of certain gases for use in ultrasound contrast which have improved persistence in the body. The gases, of which fluorine-containing gases are preferred, can be used per se (as "free gas microbubbles") or can be adopted into known carriers such as albumin microspheres, solid particles or phospholipid liposomes.

More specific support for the new claims is as follows:

Claim 15 is supported in the specification at page 15, lines 2-15.

Claims 16-18 are supported at Table II, pages 28-29 which lists perfluoropropane, perfluorobutane, perfluoropentane and sulfur hexafluoride. Perfluorohexane is supported by adding one $-CF_2-$ to perfluoro-n-pentane as disclosed at page 27, lines 7-15.

Claim 19 is supported in the specification at page 15, lines 2-15.

Claim 20 is supported in the specification at page 16, lines 16-30.

Claim 21 is supported at page 30, lines 9-20.

Claims 22-25 is supported Table II, pages 28-29 and page 27, lines 7-15. "Gas emulsion" is supported at page 32, Examples 1 and 2.

Claims 26-27 are supported at Example 1, page 32.

Claims 28 and 29 are supported at page 15, lines 34-35 through page 16, lines 1-13.

Claims 30-31 are supported at Example 2, page 32.

Claim 32 is supported at page 31, lines 3-6.

Claims 33-37 are supported at page 21, lines 20-25; page 27, lines 1-15 and Table II at pages 28-29.

Claim 38 is supported at page 12, lines 19-23.

Claim 39 is supported at page 12, lines 19-23; and page 17, line 19.

Claim 40 is supported at page 15, lines 14-33.

Claims 42-43 are supported at page 16, lines 1-7.

Claims 44-45 are supported at page 14, line 9 to page 15, line 12.

Claims 46-49 are supported at page 12, lines 19-23; page 21, lines 20-25; page 27, lines 1-15; Table II at pages 28-29 and at page 27, lines 7-15.

Claims 50-51 are supported at page 4, lines 14-25; page 19, lines 1-5; Table II at pages 28-29; at Table IV, pages 35-38 and page 27, lines 7-15.

Claim 52 is supported at page 4, lines 18-22.

Should the examiner have any questions concerning this application he is encouraged to telephone applicant's undersigned representative at 415-773-3129. Early and favorable action is respectfully requested.

Respectfully submitted,
LIMBACH & LIMBACH

Dated: Dec 20 1996

By: Patric Bengtsson
W. Patrick Bengtsson
Reg. No. 32,456
Attorneys for Applicant(s)

Atty Docket No. SNUS-124

GASEOUS ULTRASOUND CONTRAST MEDIA AND
METHOD FOR SELECTING GASES FOR USE AS
ULTRASOUND CONTRAST MEDIA

RELATED APPLICATIONS

5 This application is a continuation in part of
U.S. application Serial Number 07/893,657 filed
June 5, 1992, which is a continuation in part of U.S.
application Serial Number 07/761,311 filed
September 17, 1991.

DESCRIPTION

10 This invention relates to agents that enhance
the contrast in an ultrasound image generated for use
in medical diagnosis. The contrast-enhancing media
disclosed herein are comprised of extremely small gas
15 bubbles which are present in a solution that is
infused into the body during or just before an
ultrasound image is generated. This invention is also
directed to a method for enhancing such images by
selecting gases from which a collection of free gas
20 microbubbles can be prepared that have novel and
superior properties. These microbubbles, composed of
the gases whose selection is enabled by the process
of this invention, may be extremely small in size and
yet survive in the bloodstream long enough to allow
25 contrast-enhanced imaging of parts of the
cardiovascular system, peripheral vascular system,
and vital organs previously believed to be
inaccessible to free gas microbubbles.

BACKGROUND

30 When using ultrasound to obtain an image of the
internal organs and structures of a human or animal,
ultrasound waves -- waves of sound energy at a
frequency above that discernable by the human ear --

are reflected as they pass through the body. Different types of body tissue reflect the ultrasound waves differently and the reflections, often aptly described as "echoes," that are produced by the ultrasound waves reflecting off different internal structures are detected and converted electronically into a visual display. This display may prove invaluable to a physician or other diagnostician in several ways, including evaluating the progression of cardiovascular disease or the existence or nature of a tumor.

For some medical conditions, obtaining a useful image of the organ or structure of interest is especially difficult because the details of the structure may not be adequately discernible from the surrounding tissue in an ultrasound image produced by the reflection of ultrasound waves absent a contrast-enhancing agent. Additionally, traditional ultrasound images are notoriously poor in quality and resolution. For these reasons, detection and observation of certain physiological conditions may be substantially improved by enhancing the contrast in an ultrasound image by infusing an agent into an organ or other structure of interest. In other cases, detection of the movement of the contrast-enhancing agent itself is particularly important. For example, a distinct blood flow pattern that is known to result from particular cardiovascular abnormalities may only be discernible by infusing a contrasting agent into the bloodstream and observing the dynamics of the blood flow.

Medical researchers have made extensive investigation into the use of solids, gases and liquids in an attempt to discover ultrasound contrast-enhancing agents suitable for particular

diagnostic purposes. Composite substances such as gelatin encapsulated microbubbles, gas-incorporated liposomes, sonicated partially denatured proteins and emulsions containing highly fluorinated organic compounds have also been studied in an attempt to develop an agent that has certain ideal qualities, primarily, stability in the body and the ability to provide significantly enhanced contrast in an ultrasound image.

Small bubbles of a gas, termed "microbubbles," are readily detected in an image produced using standard ultrasound imaging techniques. When infused into the bloodstream or a particular site in the body, microbubbles enhance the contrast between the region containing the microbubbles and the surrounding tissue.

A substantial amount of the research effort directed at contrast-enhancing agents has focused on the use of extremely small gas bubbles. Investigators have long known that free gas bubbles provide a highly effective contrast agent because a gas bubble has unique physical characteristics that affect ultrasound energy as it is directed through the body. The advantages offered by free gas bubbles as opposed to liquid or solid agents that exhibit contrast enhancement is described in detail below in the context of the discussion of ultrasound diagnostic techniques.

Despite the known advantages, however, the rapid dissolution of free gas bubbles in solutions such as blood or many aqueous intravenous solutions, severely limits their use as an ultrasound contrast-enhancing agent. The most important limitations are the size of the microbubble and the length of time that a

microbubble will exist before dissolving into the solution.

Examining the size requirements for microbubbles more closely, the gas bubbles must, of course, be sufficiently small that a suspension of the bubbles does not carry the risk of embolism to the organism in which they are infused. At the same time, extremely small free gas bubbles composed of the gases generally used in ultrasound contrast imaging dissolve into solution so rapidly that their image-enhancing capability exists only immediately proximate to the infusion site. An additional obstacle exists for ultrasound imaging of the cardiovascular system. Medical researchers have studied the time required for microbubbles composed of ordinary air, pure nitrogen, pure oxygen, or carbon dioxide, to dissolve into solution. Microbubbles of these gases that are sufficiently small to be able to pass through the lungs and reach the left heart, less than about 8 microns in diameter, have a life span of less than approximately 0.25 seconds. Meltzer, R.S., Tickner, E.G., Popp, R.L., "Why Do the Lungs Clear Ultrasonic Contrast?" Ultrasound in Medicine and Biology, Vol. 6, p.263, 267 (1980). Since it takes over 2 seconds for blood to pass through the lungs, microbubbles of these gases would be fully dissolved during passage through the lungs and would never reach the left heart. Ibid. Primarily because of this tradeoff between bubble size and life span, many researchers concluded that free gas microbubbles were not useful as a contrast-enhancing agent for ultrasound diagnosis of certain parts of the cardiovascular system.

However, the ultrasound contrast-enhancing media described herein comprises microbubbles, composed of

the biocompatible gases whose selection is also provided by this invention, that are sufficiently small that they pass through the pulmonary capillary diameter of approximately 8 microns and thereby allow contrast-enhanced ultrasound diagnosis of the left chambers of the heart. The free gas microbubbles survive in the bloodstream long enough that they may be peripherally intravenously infused, travel through the right heart, through the lungs, and into the left cardiac chambers without dissolving into solution. Additionally, certain of these media have extremely long persistence in solution and will enable contrast-enhancement of many other organs and structures.

This invention overcomes many of the inherent limitations thought to exist with the use of free gas microbubbles by providing, in part, a method for selecting special gases based on particular physical criteria such that microbubbles composed of these gases do not suffer from the same limitations as the microbubbles previously investigated. Therefore, it has been discovered that the ultrasound contrast-enhancing media described herein comprising a composition of microbubbles produced using a biocompatible gas or combination of gases selected by the physical and chemical parameters disclosed herein can exist for a sufficient length of time and be of sufficiently small size that their stability in the bloodstream allows enhanced ultrasound contrast imaging of particular structures in the body previously thought inaccessible to free gas microbubbles.

By using the term "biocompatible gas" I mean a chemical entity which is capable of performing its functions within or upon a living organism in an

acceptable manner, without undue toxicity or physiological or pharmacological effects, and which is, at the temperature of the living organism, in a state of matter distinguished from the solid or liquid states by very low density and viscosity, relatively great expansion and contraction with changes in pressure and temperature, and the spontaneous tendency to become distributed uniformly throughout any container. The following Table contains the assumed body temperatures for various living organisms:

Organism	Rectal Temperature (degree Fahrenheit)
Swine (<i>Sus Scrofa</i>)	101.5-102.5
Sheep (<i>Ovis sp.</i>)	101-103
Rabbit (<i>Oryctolagus cuniculus</i>)	102-103.5
Rat (<i>Tattus morvegicus</i>)	99.5-100.6
Monkey (<i>Macaca mulatta</i>)	101-102
Mouse (<i>Mus Musculus</i>)	98-101
Goat (<i>Capra hircus</i>)	101-103
Guinea pig (<i>Cavia porcellus</i>)	102-104
Hamster (<i>Mesocricetus sp.</i>)	101-103
Ham (<i>Homo sapiens</i>)	98.6-100.4
Horse (<i>Equus sp.</i>)	101-102.5
Dog (<i>Canin familiaris</i>)	101-102
Baboon (<i>Papio</i>)	98-100
Cat (<i>Felis catus</i>)	101-102
Cattle (<i>Bos taurus</i>)	101.5-102.5
Chimpanzee (<i>Pan</i>)	96-100

Techniques For Measuring Ultrasound Contrast-Enhancement Phenomena

To more fully appreciate the subject matter of the present invention, it is useful to describe what is presently known about the technology of ultrasound imaging and to review the search for improved ultrasound contrast-enhancing agents in that light.

Materials that are useful as ultrasound contrast agents operate by having an effect on ultrasound waves as they pass through the body and are reflected

to create the image from which a medical diagnosis is made. In an attempt to develop an efficient image-contrast agent, one skilled in the art recognizes that different types of substances affect ultrasound waves in different ways and to varying degrees.

Moreover, certain of the effects caused by contrast-enhancing agents are more readily measured and observed than others. Thus, in selecting an ideal composition for a contrast-enhancing agent, one would prefer the substance that has the most dramatic effect on the ultrasound wave as it passes through the body. Also, the effect on the ultrasound wave should be easily measured. There are three main contrast-enhancing effects which can be seen in an ultrasound image: backscatter, beam attenuation, and speed of sound differential. Each of these effects will be described in turn.

A. BACKSCATTER

When an ultrasound wave that is passing through the body encounters a structure, such as an organ or other body tissue, the structure reflects a portion of the ultrasound wave. Different structures within the body reflect ultrasound energy in different ways and in varying strengths. This reflected energy is detected and used to generate an image of the structures through which the ultrasound wave has passed. The term "backscatter" refers to the phenomena in which ultrasound energy is scattered back towards the source by a substance with certain physical properties.

It has long been recognized that the contrast observed in an ultrasound image may be enhanced by the presence of substances known to cause a large amount of backscatter. When such a substance is

administered to a distinct part of the body, the contrast between the ultrasound image of this part of the body and the surrounding tissues not containing the substance is enhanced. It is well understood that, due to their physical properties, different substances cause backscatter in varying degrees. Accordingly, the search for contrast-enhancing agents has focused on substances that are stable and non-toxic and that exhibit the maximum amount of backscatter.

Making certain assumptions about the way a substance reflects ultrasound energy, mathematical formulae have been developed that describe the backscatter phenomenon. Working with these formulae, a skilled researcher can estimate the ability of gas, liquid, and solid contrast-enhancing agents to cause backscatter and the degree to which a particular substance causes measurable backscatter can be compared with other substances based on the physical characteristics known to cause the backscatter phenomenon. As a simple example, the ability of substance A to cause backscatter will be greater than substance B, if, all other factors being equal, substance A is larger than substance B. Thus, when both substances are encountered by an ultrasound wave, the larger substance scatters a greater amount of the ultrasound wave.

The capability of a substance to cause backscatter of ultrasound energy also depends on other characteristics of the substance such as its ability to be compressed. Of particular importance is the dramatic increase in backscatter caused by gas bubbles due to the bubble resonance phenomenon which is described below. When examining different substances, it is useful to compare one particular

measure of the ability of a substance to cause backscatter known as the "scattering cross-section."

The scattering cross-section of a particular substance is proportional to the radius of the scatterer, and also depends on the wavelength of the ultrasound energy and on other physical properties of the substance, J. Ophir and K. J. Parker, Contrast Agents in Diagnostic Ultrasound, Ultrasound in Medicine & Biology, vol. 15, n. 4, p. 319, 323 (1989).

The scattering cross-section of a small scatterer, a , can be determined by a known equation:

$$\sigma = \left[\frac{4}{9} \pi a^2 (ka)^4 \right] \left[\left| \frac{\kappa_s - \kappa}{\kappa} \right|^2 + \frac{1}{3} \left| \frac{3(\rho_s - \rho)}{2\rho_s - \rho} \right|^2 \right]$$

where $\kappa = 2\pi/\lambda$, where λ is the wavelength; a = the radius of the scatterer; κ_s = adiabatic compressibility of the scatterer; κ = adiabatic compressibility of the medium in which the scatterer exists, ρ_s = density of the scatterers and ρ = the density of the medium in which the scatterer exists. P. M. Morse and K. U. Ingard, Theoretical Acoustics, p. 427, McGraw Hill, New York (1968).

In evaluating the utility of different substances as image contrasting agents, one can use this equation to determine which agents will have the higher scattering cross-section and, accordingly, which agents will provide the greatest contrast in an ultrasound image.

Referring to the above equation, the first bracketed quantity in the above equation can be assumed to be constant for the purpose of comparing solid, liquid and gaseous scatterers. It can be assumed that the compressibility of a solid particle

is much less than that of the surrounding medium and that the density of the particle is much greater. Using this assumption, the scattering cross section of a solid particle contrast-enhancing agent has been
5 estimated as 1.75. Ophir and Parker, supra, at 325.

For a pure liquid scatterer, the adiabatic compressibility and density of the scatterer κ_s and the surrounding medium κ are likely to be approximately equal which would, from the above
10 equation, yield the result that liquids would have a scattering cross-section of zero. However, liquids may exhibit some backscatter if large volumes of a liquid agent are present presumably because the term a in the first bracketed quantity in the above
15 equation may become sufficiently large. For example, if a liquid agent passes from a very small vessel to a very large one such that the liquid occupies substantially all of the vessel the liquid may exhibit measurable backscatter. Nevertheless, in
20 light of the above equation and the following, it is appreciated by those skilled in the art that pure liquids are relatively inefficient scatterers compared to free gas microbubbles.

It is known that changes in the acoustic
25 properties of a substance are pronounced at the interface between two phases, i.e., liquid/gas, because the reflection characteristics of an ultrasound wave change markedly at this interface. Additionally, the scatter cross-section of a gas is
30 substantially different than a liquid or solid, in part, because a gas bubble can be compressed to a much greater degree than a liquid or solid. The physical characteristics of gas bubbles in solution are known and standard values for compressibility and
35 density figures for ordinary air can be used in the

above equation. Using these standard values, the result for the second bracketed term alone in the above equation is approximately 10^{14} , Ophir and Parker supra, at 325, with the total scattering cross section varying as the radius a of the bubble varies. Moreover, free gas bubbles in a liquid exhibit oscillatory motion such that, at certain frequencies, gas bubbles will resonate at a frequency near that of the ultrasound waves commonly used in medical imaging. As a result, the scattering cross-section of a gas bubble can be over a thousand times larger than its physical size.

Therefore, it is recognized that gas micro-bubbles are superior scatterers of ultrasound energy and would be an ideal contrast-enhancing agent if the obstacle of their rapid dissolution into solution could be overcome.

B. BEAM ATTENUATION

Another effect which can be observed from the presence of certain solid contrast-enhancing agents, is the attenuation of the ultrasound wave. Image contrast has been observed in conventional imaging due to localized attenuation differences between certain tissue types. K. J. Parker and R. C. Wang, "Measurement of Ultrasonic Attenuation Within Regions selected from B-Scan Images," IEEE Trans. Biomed. Enar. BME 30(8), p. 431-37 (1983); K. J. Parker, R. C. Wang, and R. M. Lerner, "Attenuation of Ultrasound Magnitude and Frequency Dependence for Tissue Characterization," Radiology, 153(3), p. 785-88 (1984). It has been hypothesized that measurements of the attenuation of a region of tissue taken before and after infusion of an agent may yield an enhanced image. However, techniques based on attenuation contrast as a means to measure the contrast

enhancement of a liquid agent are not well-developed and, even if fully developed, may suffer from limitations as to the internal organs or structures with which this technique can be used. For example, it is unlikely that a loss of attenuation due to liquid contrast agents could be observed in the image of the cardiovascular system because of the high volume of liquid contrast agent that would need to be present in a given vessel before a substantial difference in attenuation could be measured.

Measurement of the attenuation contrast caused by microspheres of Albunex (Molecular Biosystems, San Diego, CA) in vitro has been accomplished and it has been suggested that in vivo attenuation contrast measurement could be achieved. H. Bleeker, K. Shung, J. Burnhart, "On the Application of Ultrasonic Contrast Agents for Blood Flowometry and Assessment of Cardiac Perfusion," J. Ultrasound Med. 9:461-471 (1990). Albunex is a suspension of 2-4 micron encapsulated air-filled microspheres that have been observed to have acceptable stability in vivo and are sufficiently small in size that contrast enhancement can occur in the left atrium or ventricle. Also, attenuation contrast resulting from iodipamide ethyl ester (IDE) particles accumulated in the liver has been observed. Under such circumstances, the contrast enhancement is believed to result from attenuation of the ultrasound wave resulting from the presence of dense particles in a soft medium. The absorption of energy by the particles occurs by a mechanism referred to as "relative motion." The change in attenuation caused by relative motion can be shown to increase linearly with particle concentration and as the square of the density difference between the particles and the surrounding

medium. K. J. Parker, et al., "A Particulate Contrast Agent with Potential for Ultrasound Imaging of Liver," Ultrasound in Medicine & Biology, Vol. 13, No. 9, p. 555, 561 (1987). Therefore, where
5 substantial accumulation of solid particles occurs, attenuation contrast may be a viable mechanism for observing image contrast enhancement although the effect is of much smaller magnitude than the backscatter phenomenon and would appear to be of
10 little use in cardiovascular diagnoses.

C. SPEED OF SOUND DIFFERENTIAL

An additional possible technique to enhance contrast in an ultrasound image has been proposed based on the fact that the speed of sound varies
15 depending on the media through which it travels. Therefore, if a large enough volume of an agent, through which the speed of sound is different than the surrounding tissue, can be infused into a target area, the difference in the speed of sound through
20 the target area may be measurable. Presently, this technique is only experimental.

Therefore, considering the three techniques described above for contrast enhancement in an ultrasound image, the marked increase in backscatter
25 caused by free gas microbubbles is the most dramatic effect and contrast-enhancing agents that take advantage of this phenomenon would be the most desirable if the obstacle of their limited stability in solution could be overcome.

30 The Materials Presently Used
as Contrast-Enhancing Agents

In light of what is known about the various techniques described above, attempts to develop a contrast-enhancing agent whose presence generates

substantial contrast in an ultrasound image, and whose survival in vivo is sufficiently long to allow contrast-enhanced imaging of the cardiovascular system, has led to the investigation of a broad variety of substances -- gases, liquids, solids, and combinations of these -- as potential contrast-enhancing agents.

A. SOLID PARTICLES

Typically, the solid substances that have been studied as potential contrast-enhancing agents are extremely small particles that are manufactured in uniform size. Large numbers of these particles can be infused and circulate freely in the bloodstream or they may be injected into a particular structure or region in the body.

IDE particles are solid particles that can be produced in large quantities with a relatively narrow size distribution of approximately 0.5-2.0 microns. Sterile saline injections of these particles may be injected and will tend to accumulate in the liver. Once a substantial accumulation occurs, contrast enhancement may be exhibited by either attenuation contrast or backscatter mechanisms. Although suspensions comprising these solid particles dispersed in a liquid may exhibit acceptable stability, the backscatter or attenuation effects are relatively minor compared to free gas bubbles and a substantial accumulation of the particles must occur before appreciable contrast is observed in an ultrasound image. Thus, use of these suspensions has been limited to certain cell types in which the particles have the tendency to coagulate because unless the suspension becomes highly concentrated in particular tissue, the contrast enhancement will be minor.

SHU-454 (Schering, A. G., West Berlin, Germany) is an experimental contrast-enhancing agent in powder form that, when mixed with a saccharide diluent, forms a suspension of crystals of various rhomboid and polyhedral shapes ranging in size from 5 to 10 microns. Although the precise mechanism by which these crystals enhance ultrasound contrast is not completely understood, it is suspected that the crystals may trap microbubbles in their structure or that the crystals themselves may backscatter ultrasound energy by an as-yet undetermined mechanism.

B. LIQUIDS AND EMULSIONS

In another attempt to achieve a satisfactory agent, emulsions are prepared by combining a chemical species compatible with body tissue and a species that provides high ultrasound contrast enhancement. European Patent Application 0231091 discloses emulsions of oil in water containing highly fluorinated organic compounds that have been studied in connection with their possible use as a blood substitute and are also capable of providing enhanced contrast in an ultrasound image.

Emulsions containing perfluorooctyl bromide (PFOB) have also been examined. Perfluorooctyl bromide emulsions are liquid compounds known to have the ability to transport oxygen. PFOB emulsions have exhibited a limited utility as ultrasound contrast agents because of a tendency to accumulate in certain types of cells. Although the mechanism is not completely understood, PFOB emulsions may provide ultrasound contrast because of their high density and relatively large compressibility constant.

United States Patent No. 4,900,540 describes the use of phospholipid-based liposomes containing a gas

or gas precursor as a contrast-enhancing agent. A liposome is a microscopic, spherical vesicle, containing a bilayer of phospholipids and other amphipathic molecules and an inner aqueous cavity, all of which is compatible with the cells of the body. In most applications, liposomes are used as a means to encapsulate biologically active materials. The above reference discloses the use of a gas or gas precursors incorporated into the liposome core to provide a longer life span for the gas when infused into the body. Production of stable liposomes is an expensive and time consuming process requiring specialized raw materials and equipment.

C. MICROBUBBLES

As noted above, a critical parameter that must be satisfied by a microbubble used as a contrast-enhancing agent is size. Free gas microbubbles larger than approximately 8 microns may still be small enough to avoid impeding blood flow or occluding vascular beds. However, microbubbles larger than 8 microns are removed from the bloodstream when blood flows through the lungs. As noted above, medical researchers have reported in the medical literature that microbubbles small enough to pass through the lungs will dissolve so quickly that contrast enhancement of left heart images is not possible with a free gas microbubble. Meltzer, R.S., Tickner, E.G., Popp, R.L., "Why Do the Lungs Clear Ultrasonic Contrast?" Ultrasound in Medicine and Biology, vol. 6, p.263, 267 (1980).

However, cognizant of the advantages to be gained by use of microbubbles as contrast-enhancing agents by virtue of their large scattering cross-section, considerable attention has been focused on developing mixtures containing microbubbles that are

rendered stable in solution. Enhancing the stability of gas microbubbles may be accomplished by a number of techniques.

Each of the following techniques essentially involves suspending a collection of microbubbles in a substrate in which a bubble of ordinary gas is more stable than in the bloodstream.

In one approach, microbubbles are created in viscous liquids that are injected or infused into the body while the ultrasound diagnosis is in progress. The theory behind the use of viscous fluids involves an attempt to reduce the rate at which the gas dissolves into the liquid and, in so doing, provide a more stable chemical environment for the bubbles so that their lifetime is extended.

Several variations on this general approach have been described. EPO Application No. 0324938 describes a viscous solution of a biocompatible material, such as a human protein, in which microbubbles are contained. By submitting a viscous protein solution to sonication, microbubbles are formed in the solution. Partial denaturation of the protein by chemical treatment or heat provides additional stability to microbubbles in the solution by decreasing the surface tension between bubble and solution.

Therefore, the above approaches may be classified as an attempt to enhance the stability of microbubbles by use of a stabilizing medium in which the microbubbles are contained. However, none of these approaches have addressed the primary physical and chemical properties of gases which have seriously limited the use of free gas microbubbles in ultrasound diagnosis, particularly with respect to the cardiovascular system. None of these approaches

suggest that selection of the gases, by precise criteria, would yield the ability to produce stable microbubbles at a size that would allow transpulmonary contrast-enhanced ultrasound imaging.

5 The behavior of microbubbles in solution can be described mathematically based on certain parameters and characteristics of the gas of which the bubble is formed and the solution in which the bubble is present. Depending on the degree to which a solution
10 is saturated with the gas of which the microbubbles are formed, the survival time of the microbubbles can be calculated. P.S. Epstein, M.S. Plesset, "On the Stability of Gas Bubbles in Liquid-Gas Solutions,"
15 The Journal of Chemical Physics, Vol. 18, n. 11, 1505 (1950). Based on calculations, it is apparent that as the size of the bubble decreases, the surface tension between bubble and surrounding solution increases. As the surface tension increases, the rate at which the bubble dissolves into the solution
20 increases rapidly and, therefore, the size of the bubble decreases more and more rapidly. Thus, the rate at which the bubble shrinks increases as the size of the bubble decreases. The ultimate effect of this is that a population of small free gas
25 microbubbles composed of ordinary air dissolves so rapidly that the contrast-enhancing effect is extremely short lived. Using known mathematical formula, one can calculate that a microbubble of air that is 8 microns in diameter, which is small enough
30 to pass through the lungs, will dissolve in between 190 and 550 milliseconds depending on the degree of saturation of the surrounding solution. Based on these calculations, medical investigators studying the way in which the lungs remove ultrasound contrast
35 agent have calculated the dissolution times of oxygen

and nitrogen gas microbubbles in human and canine blood and have concluded that free gas microbubble contrast agents will not allow contrast-enhanced imaging of the left ventricle because of the extremely brief life of the microbubbles.

The physical properties of the systems that feature gas bubbles or gases dissolved in liquid solutions have been investigated in detail including the diffusion of air bubbles formed in the cavitating flow of a liquid and the scatter of light and sound in water by gas bubbles.

The stability of gas bubbles in liquid-gas solution has been investigated both theoretically, Epstein P.S. and Plesset M.S., On the Stability of Gas Bubbles in Liquid-Gas Solutions, J. Chem. Phys. 18:1505-1509 (1950) and experimentally, Yang WJ, Dynamics of Gas Bubbles in Whole Blood and Plasma, J. Biomech 4:119-125 (1971); Yang WJ, Echigo R., Wotton DR, and Hwang JB, Experimental Studies of the Dissolution of Gas Bubbles in Whole Blood and Plasma-I. Stationary Bubbles. J. Biomech 3:275-281 (1971); Yang WJ, Echigo R., Wotton DR, Hwang JB, Experimental Studies of the Dissolution of Gas Bubbles in Whole Blood and Plasma-II. Moving Bubbles or Liquids. J. Biomech 4:283-288 (1971). The physical and chemical properties of the liquid and the gas determine the kinetic and thermodynamic behavior of the system. The chemical properties of the system which influence the stability of a bubble, and accordingly the life time, are the rate and extent of reactions which either consume, transform, or generate gas molecules.

For example, a well known reaction that is observed between a gas and a liquid takes place when carbon dioxide gas is present in water. As the gas dissolves into the aqueous solution, carbonic acid is

created by hydration of the carbon dioxide gas. Because carbon dioxide gas is highly soluble in water, the gas diffuses rapidly into the solution and the bubble size diminishes rapidly. The presence of the carbonic acid in the solution alters the acid-base chemistry of the aqueous solution and, as the chemical properties of the solution are changed by dissolution of the gas, the stability of the carbon dioxide gas bubbles changes as the solution becomes saturated. In this system, the rate of dissolution of a gas bubble depends in part on the concentration of carbon dioxide gas that is already dissolved in solution.

However, depending on the particular gas or liquid present in the system, the gas may be substantially insoluble in the liquid and dissolution of a gas bubble will be slower. In this situation, it has been discovered that it is possible to calculate bubble stability in a gas-liquid system by examining certain physical parameters of the gas.

BRIEF DESCRIPTION OF THE INVENTION

It has been discovered that it is possible to identify chemical systems where extremely small gas bubbles are not reactive in an aqueous solution. Relying on the method disclosed herein one skilled in the art may specially select particular gases based on their physical and chemical properties for use in ultrasound imaging. These gases can be used to produce the contrast-enhancing media that is also the subject matter of this invention. The microbubbles can be produced using certain existing techniques that use ordinary air, and can be infused as in a conventional ultrasound diagnosis.

5 The method that is the subject matter of this invention requires that calculations be made, consistent with the equations provided herein, based on the intrinsic physical properties of a gas and a liquid. Particularly, the density of a gas, the solubility of a gas in solution, and the diffusivity of a gas in solution, which in turn is dependent on the molar volume of the gas and the viscosity of the solution, are used in the equations disclosed below. Thus, by the method disclosed herein, the physical properties of a given gas-liquid system can be evaluated, the rate and extent of bubble collapse can be estimated, and gases that would constitute effective contrast-enhancing agents can be selected based on these calculations. Using existing techniques, substantially improved contrast-enhancing media may then be produced and used to improve the quality and usefulness of ultrasound imaging.

DETAILED DESCRIPTION OF THE INVENTION

20 To understand the method of this invention, it is useful to derive the mathematical relationships that describe the parameters of a gas-liquid system and the effect on bubble stability that occurs when a value for one or more of these parameters is altered. It is assumed that, at an initial time, T_0 , a spherical gas bubble of gas X, with a radius of R_0 , is placed in a solution in which the initial concentration of gas X dissolved in the solution is equal to zero. Over some period of time, the bubble of gas X will dissolve into the solvent at which time its radius R will equal zero. Assume further that the solution is at constant temperature and pressure and that the dissolved gas concentration for a solution saturated with the particular gas is

designated C_s . Thus, at T_0 , the concentration of the gas in the solution is zero, meaning that none of the gas has yet dissolved and all of the gas that is present is still contained within the bubble of radius R_0 .

As time progresses, due to the difference in the concentration of the gas in the bubble and the gas in solution, the bubble will tend to shrink as gas in the bubble is dissolved into the liquid by the process of diffusion. The change in bubble radius from its original radius of R_0 to, after the passage of a particular amount of time, a smaller radius R is expressed by Equation (1),

$$\frac{R}{R_0} = \left[1 - \left(\frac{2DC_s}{\rho R_0^2} \right) T \right]^{1/2}$$

where R is the bubble radius at time T , D is the coefficient of diffusivity of the particular gas in the liquid, and ρ is the density of the particular gas of which the bubble is composed.

It follows that the time T required for a bubble to dissolve completely may be determined from Equation (1) by setting $R/R_0 = 0$, and solving for T :

$$\text{Equation (2)} \quad T = \frac{R_0^2 \rho}{2DC_s}$$

This result qualitatively indicates that bubble stability, and hence life span, is enhanced by increasing the initial bubble size R_0 or by selecting a gas of higher density ρ , lower solubility C_s in the liquid phase, or a lower coefficient of diffusivity D .

The diffusivity D of a gas in a liquid is dependent on the molar volume of the gas (V_m), and

the viscosity of the solution (η) as expressed by a known Equation;

$$\text{(Equation 3)} \quad D = 13.26 \times 10^{-5} \cdot \eta^{-1.14} \cdot V_m^{-0.589}$$

By substituting the expression for D given in Equation (3) into Equation (2) it is revealed that bubble stability is enhanced by using gases of larger molar volume V_m , which tend to have a higher molecular weight, and liquids of higher viscosity.

By way of example, a comparison of the stability of air microbubbles and microbubbles composed of gases specially selected by the method disclosed herein may be made. Taking the value of D for air in water at 22°C as $2 \times 10^{-5} \text{ cm}^2 \text{ sec}^{-1}$ and the ratio $C_s/\rho = 0.02$ (Epstein and Plesset, Ibid.), one obtains the following data for the time t for complete solution of air bubbles in water (unsaturated with air) :

TABLE I

	INITIAL BUBBLE DIAMETER, microns	TIME, milliseconds
20	12	450
	10	313
	8	200
	6	113
	5	78
25	4	50
	3	28
	2	13
	1	3

If the blood transit time from the pulmonary capillaries to the left ventricle is two seconds or more (Hamilton, W.F. editor, Handbook of Physiology, Vol. 2, section 2, CIRCULATION. American Physiology Society, Washington, D.C., p. 709, (1963)), and recognizing that only microbubbles of approximately 8 microns or less will be small enough to pass through the lungs, it is clear that none of these bubbles

have a life span in solution long enough to be useful contrast agents for ultrasound contrast-enhanced imaging of the left ventricle.

5 The method of the present invention allows identification of potentially useful gases by comparing the properties of any particular gas, denoted gas X in the following description, to air. Taking Equations (2) and (3) above, a coefficient Q may be formulated for a particular gas X that will describe the stability of microbubbles composed of gas X in a given liquid. The value of the Q coefficient determined by this method for a particular gas X also can be used to determine the utility of gas X as an ultrasound contrast-enhancing agent as compared to ordinary air.

15 From Equation (2) above, an equation that describes the time for complete dissolution of a bubble of gas X compared to the same size bubble of ordinary air under identical conditions of solution temperature and solution viscosity may be written based on the physical properties of gas X and air: Equation (4)

$$25 \quad T_x = T_{\text{air}} \left[\frac{\rho_x}{\rho_{\text{air}}} \right] \left[\frac{(C_s)_{\text{air}}}{(C_s)_x} \right] \left[\frac{D_{\text{air}}}{13.26 \times 10^{-5} \cdot \eta^{-1.14} \cdot (V_m)_x^{-.589}} \right]$$

or, if D is known for gas X,
Equation (5)

$$30 \quad T_x = T_{\text{air}} \left[\frac{\rho_x}{\rho_{\text{air}}} \right] \left[\frac{(C_s)_{\text{air}}}{(C_s)_x} \right] \left[\frac{D_{\text{air}}}{D_x} \right]$$

To formulate this equation so that the value Q may be obtained to enable comparison of gas X with air, the above equation may be rewritten:

Equation (6) $T_x = QT_{air}$ where

$$Q = \left[\frac{\rho_x}{\rho_{air}} \right] \left[\frac{(C_s)_{air}}{(C_s)_x} \right] \left[\frac{D_{air}}{D_x} \right]$$

Assuming for comparison, a solution of water at 22 degrees C, the density, diffusivity, and solubility of air in the solution are known quantities which may be substituted into the above equation yielding:

$$\text{Equation (7)} \quad Q = 4.0 \times 10^{-7} \left[\frac{\rho_x}{(C_s)_x D_x} \right]$$

Substituting Equation (3) into the above for gases whose diffusivity D_x is not readily known, and assuming that the viscosity term η below for water at 22 degrees C is approximately equal to 1.0 cP,

$$\text{Equation (8)} \quad Q = 3.0 \times 10^{-3} \left[\frac{\rho_x}{(C_s)_x (V_m)_x^{-.589}} \right]$$

Thus, knowing the density, solubility and molar volume of a gas, this method allows the calculation of the value of the Q coefficient.

If Q is less than one, microbubbles of gas X will be less stable in a given solvent than microbubbles of air. If Q is greater than one, microbubbles formed of gas X are more stable than microbubbles of air and will survive in solution longer than air bubbles. All other properties being the same for a given microbubble size, the time for complete dissolution of a microbubble of gas X is equal to the time for complete dissolution of a microbubble of ordinary air multiplied by the Q

coefficient. For example, if the Q coefficient for gas X is 10,000, a microbubble of gas X will survive 10,000 times as long in solution compared to a microbubble of air. A Q value can be determined for any gas in any solution assuming the quantities identified herein are known or can be estimated.

Different methods for determining or estimating values for the individual parameters of density, diffusivity, and solubility may be needed depending on the chemical structure of the gas. Values for these parameters may or may not be available from known scientific literature sources such as the *Gas Encyclopedia* or the tabulations published by the American Chemical Society. Values for the density of most gases are readily available from sources such as the Handbook of Chemistry and Physics, CRC Press, 72d Ed. (1991-92). Additionally, the solubility in water and molar volume of some gases has been measured with accuracy. In many cases however, calculations for the numerical values for molar volume and solubility may need to be calculated or estimated to provide the data used to determine the value of the Q coefficient for an individual gas by the method described above. An example of the calculation of Q values for a preferred selection of gases illustrates how the method of this invention can be applied to individual gases.

Generally, many fluorine-containing gases exhibit extremely low solubility in water, and have relatively high molecular weights, high molar volumes, and high densities. To determine the Q value for several gases, the solubility, molar volume and density of the individual gases are determined and the values are substituted into Equations (7) or (8) above.

Determination of Gas Solubility for Fluorocarbons

This method for estimating the gas solubility of fluorocarbons uses extrapolation of the experimental data of Kabalnov AS, Makarov KN, and Scherbakova OV.

5 "Solubility of Fluorocarbons in Water as a Key
Parameter Determining Fluorocarbon Emulsion
Stability," J. Fluor. Chem. 50, 271-284, (1990). The
gas solubility of these fluorocarbons is determined
relative to perfluoro-n-pentane which has a water
10 solubility of 4.0×10^{-6} moles per liter. For a
homologous series of non-branched fluorocarbons, the
gas solubility may be estimated by increasing or
reducing this value by a factor of about 8.0 for each
increase or reduction in the number of additional
15 $-CF_2-$ groups present in the molecule.

Determination of Molar Volume

The molar volume (V_m) is estimated from the data of Bondi A., "Van der Waals Volumes and Radii," J. Phys. Chem. 68, 441-451 (1964). The molar volume of
20 a gas can be estimated by identifying the number and
type of atoms that make up the molecule of gas in
question. By determining the number and type of
atoms present in the molecule and how the individual
atoms are bound to each other, known values may be
25 applied for the molecular volume of the individual
atoms. By considering the contribution of each
individual atom and its frequency of occurrence, one
may calculate the total molar volume for a particular
gas molecule. This calculation is best demonstrated
30 with an example.

It is known that a carbon molecule in an alkane
carbon-carbon bond has a molar volume of 3.3 cubic
centimeters per mole, a carbon atom in an alkene

carbon-carbon bond has a molar volume of 10.0 cubic centimeters per mole, and when multiple fluorine atoms are bound to an alkane carbon, a fluorine atom has a molar volume of 6.0 cubic centimeters per mole.

5 Examining octafluoropropane, this molecule contains three carbon atoms in alkane carbon-carbon bonds (3 atoms at 3.3 cubic centimeters per mole) and 6 fluorine atoms bound to alkane carbons (6 atoms at 6.0 cubic centimeters per mole) hence, octafluoro-
10 propane has a molar density of 58 cubic centimeters per mole.

Once density, molar volume, and solubility are determined, the Q value is calculated using Equation 8 above.

15 The following Table lists the Q value for a number of gases based on the calculations detailed above.

TABLE II

20	GAS	DENSITY	SOLUBILITY	MOLAR VOLUME	Q
		kg/m3	micromoles/ liter	cm3/mole	
	Argon	1.78	1500	17.9	20
	n-Butane	2.05	6696	116	5
25	Carbon Dioxide	1.98	33000	19.7	1
	Decafluorobutane	11.21	32	73	13,154
	Dodecafluoropentane	12.86	4	183	207,437
	Ethane	1.05	2900	67	13
	Ethyl ether	2.55	977,058	103	0.1
30	Helium	0.18	388	8	5
	Hexafluorobuta-1,3-diene	9 (*)	2000	56	145
	Hexafluoro-2-butyne	9 (*)	2000	58	148
	Hexafluoroethane	8.86	2100	43	116
	Hexafluoropropane	10.3	260	58	1299
35	Krypton	3.8	2067	35	44
	Neon	0.90	434	17	33
	Nitrogen	##	##	##	1
	Octafluoro-2-butene	10 (*)	220	65	1594
	Octafluorocyclobutane	9.97	220	61	1531
40	Octafluoropropane	10.3	260	58	1299
	Pentane	2	1674	113	58

Propane	2.02	2902	90	30
Sulfur Hexafluoride	5.48	220	47	722
Xenon	5.90	3448	18	28

* These density values are estimated from the known density of homologous fluorocarbons.

The solubility/density ratio value of 0.02 (supra) and the diffusivity of $2 \times 10^{-5} \text{ cm}^2 \text{ sec}^{-1}$ given above were used in equation 7 for this Q-value determination.

Once the Q value has been determined the utility of an individual gas as an ultrasound contrast-enhancing agent can be analyzed by determining the life span of a collection of microbubbles composed of the gas in question at different sizes, as was done for air in Table I above. Taking the value of Q for decafluorobutane and examining the time necessary for various sized bubbles to dissolve in water, one obtains the values in Table III below by multiplying each of the time values in Table I by the Q value for decafluorobutane:

TABLE III

INITIAL BUBBLE DIAMETER, microns	TIME, minutes
12	99
10	69
8	44
6	25
5	17
4	11
3	6.1
2	2.9
1	0.7

Notice that the time scale in Table III is minutes rather than milliseconds as was the case for air. All bubbles of decafluorobutane, even as small as 1 micron, can be injected peripherally and will not dissolve into solution during the approximately 10 seconds needed to reach the left ventricle. Similar calculations can be performed for a gas with any Q coefficient. Slightly larger bubbles will be

able to pass through the lungs and yet survive long enough to permit both examination of myocardial perfusion and dynamic abdominal organ imaging. Moreover, as with many of the gases identified by this method, decafluorobutane features low toxicity at small dosages and would, therefore, offer substantial advantages as a contrast-enhancing agent in conventional ultrasound diagnosis.

Manual creation of a microbubble suspension may be accomplished by several methods. United States Patent 4,832,941, the disclosure of which is incorporated herein by reference, refers to a method for producing a suspension of microbubbles with a diameter less than seven microns created by spraying a liquid through a quantity of gas using a three-way tap. Although techniques could vary in practice, the three-way tap is a preferred method to manually suspend a quantity of high Q coefficient gas to produce the contrast-enhancing media described herein.

The general techniques for use of a three-way tap device are well known in connection with preparation of the common Freund's adjuvant for immunizing research animals. Typically, a three-way tap is comprised of a pair of syringes, both of which are connected to a chamber. The chamber has outlet from which the suspension may be collected or infused directly.

Techniques for use of the three-way tap may differ from that described in U.S. Patent No. 4,832,941 because different gases are being used in this procedure. For example, use of one of the high Q coefficient gases disclosed herein may be more efficient if the system is purged of ordinary air or

flushed with another gas before the microbubble suspension is produced.

In a preferred embodiment of the present invention, a 40-50% Sorbitol (D-glucitol) solution is
5 mixed with approximately 1-10% by volume of a high Q-coefficient gas with approximately 5% gas being an optimal value. Sorbitol is a commercially available compound that when mixed in an aqueous solution substantially increases the viscosity of the
10 solution. Higher viscosity solutions, as seen in equation 3 above, extend the life of a microbubble in solution. A 40-50% Sorbitol solution is preferred to maintain as a bolus upon injection; that is as intact as possible without exceeding a tolerable injection
15 pressure. To produce the suspension of microbubbles, a quantity of the chosen gas is collected in a syringe. In the same syringe, a volume of the Sorbitol solution may be contained. A quantity of Sorbitol solution is drawn into the other syringe so
20 that the sum of the two volumes yields the proper percentage of gas based on the volume percentage of microbubbles desired. Using the two syringes, each featuring a very small aperture, the liquid is sprayed into the gas atmosphere approximately 25
25 times or as many times as is necessary to create a suspension of microbubbles whose size distribution is acceptable for the purposes described herein. This technique may be varied slightly, of course, in any manner that achieves the resulting suspension of
30 microbubbles of the desired size in a desired concentration. Microbubble size may be checked either visually or electronically using a Coulter Counter (Coulter Electronics) by a known method, Butler, B.D., "Production of Microbubble for Use as

Echo Contrast Agents," J. Clin. Ultrasound, V.14 408 (1986).

EXAMPLES

5 **EXAMPLE 1.** An ultrasound contrast agent was prepared using decafluorobutane as the microbubble-forming gas. A solution was prepared containing:

	sorbitol	20.0 g
	NaCl	0.9 g
	soy bean oil	6.0 mL
10	Tween 20	0.5 mL
	water q.s.	100.0 mL

15 A soapy, clear, yellow solution was afforded with stirring. A 10 mL aliquot of this solution was taken up in a 10 mL glass syringe. The syringe was then attached to a three-way stopcock. A second 10 mL syringe was attached to the stopcock and 1.0 cc of decafluorobutane (PCR, Inc., Gainesville, FL) was delivered to the empty syringe. The stopcock valve was opened to the solution-containing syringe and the liquid and gas phases mixed rapidly 20-30 times. A resulting milky-white, slightly viscous solution was obtained.

25 **EXAMPLE 2.** The gas emulsion obtained in Example 1 was diluted with water (1:10 to 1:1000), placed in a hemocytometer, and examined under the microscope using an oil immersion lens. The emulsion consisted of predominately 2-5 micron bubbles. The density was 50-100 million microbubbles per mL of original undiluted formulation.

30 **EXAMPLE 3.** The formulation of Example 1 was prepared and echocardiography performed in a canine model. A 17.5 kg mongrel dog was anesthetized with isoflurane and monitors established to measure ECG, blood

pressure, heart rate, and arterial blood gases according to the methods described by Keller, MW, Feinstein, SB, Watson, DD: Successful left ventricular opacification following peripheral venous injection of sonicated contrast agent: An experimental evaluation. Am Heart J 114:570d (1987).

The results of the safety evaluation are as follows:

**MAXIMUM PERCENTAGE CHANGE IN MEASURED
PARAMETER WITHIN 5 MIN POST INJECTION**

10

DOSE	AORTIC PRESSURE			BLOOD GASES			HEART RATE	
	SYSTOLIC	DIASTOLIC	MEAN	PaO2	PaCO2	pH		
	mm Hg							
0.5 mL	+6, -14	+9, 0	+8, -6	329	58.1	7.26	+10	-19
1.0 mL	+9, -2	+5, -1	+4, -4				+1,	-4
2.0 mL	+5, -3	+5, -1	+5, -1				0,	-1
3.0 mL	+6, -2	+7, 0	+4, -3				0,	-3
4.0 mL	+5, -1	+3, -3	+5, -3				0,	-3
5.0 mL	0, -10	+1, -3	0, -4				+1,	-1
7.0 mL	0, -13	0, -8	0, -9	313	28.6	7.36	0,	-1

20

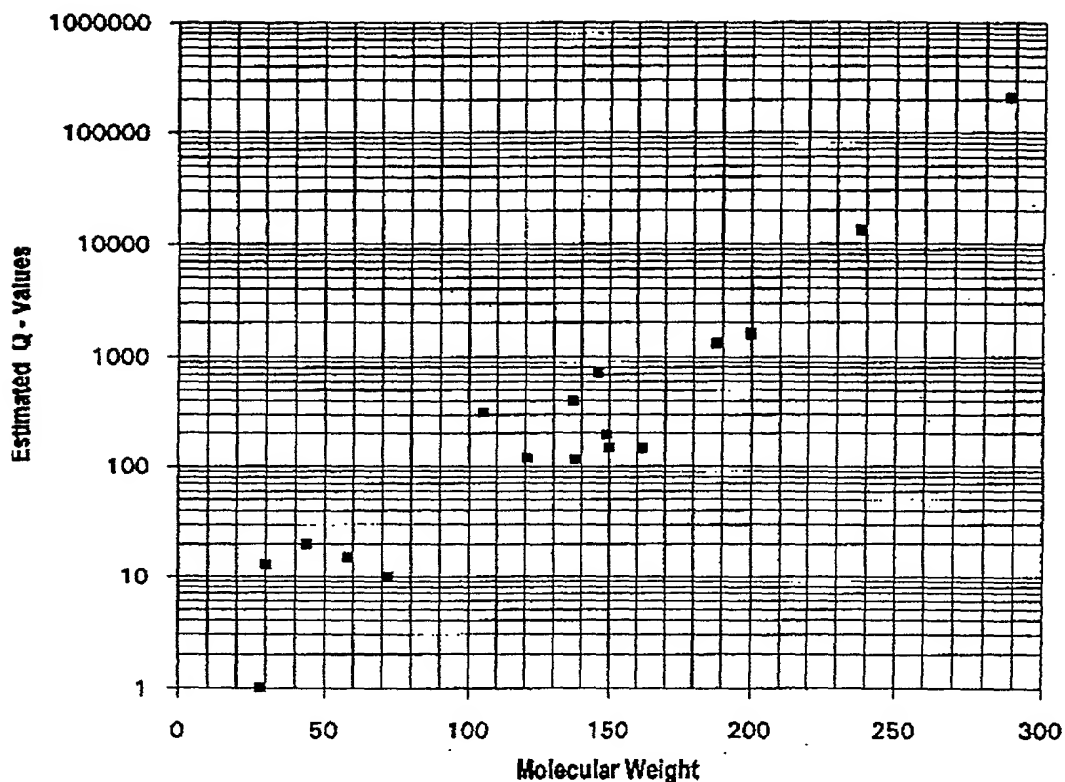
All changes were transient and returned to baseline values typically within 3-6 minutes. The above safety data demonstrate minimal changes in the measured hemodynamic parameter. All doses provided both right and left ventricular chamber opacification. The intensity increased with the increasing dose.

25

Example 4. The above specific determinations of the suitability of a particular gas for use as an ultrasound agent can be approximated if the molecular weight of a particular gas is known, can be calculated, or can be measured. This approximation is based on the determination that there is a linear relationship

30

between the logarithm of the Q-value and the molecular weight for a gas, as shown in the Figure below.



Based on this Figure, the following guidelines can be used to estimate a Q-value:

5	Molecular Weight	Estimated Q-Value
	< 35	< 5
	35-70	5-20
	71-100	21-80
	101-170	81-1000
10	171-220	1001-10,000
	221-270	10,001-100,000
	>270	>100,000

The following Table contains a series of gases with the relevant data on molecular weight and estimated Q-value. The higher the Q-value the more

promising is the particular gas. Especially promising are gases with Q-values greater than five. Additional issues, including, but not limited to, cost and toxicity, should be considered in addition to longevity of the derived microbubbles (as estimated by the Q-value) in determining the suitability of any particular gas as an ultrasound contrast agent.

TABLE IV

	<u>Chemical Name</u>	<u>Molecular Weight</u>	<u>Estimated Q Value</u>
10	Acetone, hexafluoro	166.02	81-1000
	Acetylene, isopropyl	68	5-20
	Air	28.4	<5
	Allene	40.06	5-20
15	Allene, tetrafluoro	112.03	81-1000
	Argon	39.98	5-20
	Borne, dimethyl, methoxy	71.19	21-80
	Borne, trimethyl	55.91	5-20
	Boron fluoride dihydrate	103.84	81-1000
20	1,2-Butadiene	54.09	5-20
	1,3-Butadiene	54.09	5-20
	1,3-Butadiene, 1,2,3-trichloro	157.43	81-1000
	1,3-Butadiene, 2-fluoro	72.08	21-80
	1,3-Butadiene, 2-methyl	68.12	5-20
25	1,3-Butadiene, hexafluoro	162.03	81-1000
	Butadiyne	50.06	5-20
	n-Butane	58.12	5-20
	Butane, 1-fluoro	76.11	21-80
	Butane, 2-methyl	72.15	21-80
30	Butane, decafluoro	238.03	10,001-100,000
	1-Butene	56.11	5-20
	2-Butene {cis}	56.11	5-20
	2-Butene {trans}	56.11	5-20
	1-Butene, 2-methyl	70.13	5-20
35	1-Butene, 3-methyl	70.13	5-20
	2-Butene, 3-methyl	68	5-20
	1-Butene, perfluoro	200.03	1001-10,000
	2-Butene, perfluoro	200.03	1001-10,000
	3-Butene-2-one, 4-phenyl {trans}	146.19	81-1000
40	1-Butene-3-yne, 2-methyl	66.1	5-20
	Butyl nitrite	103.12	81-100
	1-Butyne	54.09	5-20
	2-Butyne	54.09	5-20
	Butyne, 2-chloro-1,1,1,4,4,4-hexafluoro	199	1001-10,000
45	1-Butyne, 3-methyl	68.12	5-20
	2-Butyne, perfluoro	162.03	81-1000
	Butyraldehyde, 2-bromo	151	81-1000
	Carbon dioxide	44.01	5-20
	Carbonyl sulfide	60.08	5-20
50	Crotononitrile	67.09	5-20

	Cyclobutane	56.11	5-20
	Cyclobutane, methyl	70.13	5-20
	Cyclobutane, octafluoro	200.03	1001-100,000
	Cyclobutene, perfluoro	162.03	81-1000
5	Cyclopentene, 3-chloro	102.56	81-1000
	Cyclopropane	42.08	5-20
	Cyclopropane, 1,2-dimethyl {trans, dl}	70.13	5-20
	Cyclopropane, 1,1-dimethyl	70.13	5-20
	Cyclopropane, 1,2-dimethyl {cis}	70.13	5-20
10	Cyclopropane, 1,2-dimethyl {trans, l}	70.13	5-20
	Cyclopropane, ethyl	70.13	5-20
	Cyclopropane, methyl	56.11	5-20
	Deuterium	4.02	<5
	Diacetylene	50.08	5-20
15	Diaziridine, 3-ethyl-3-methyl	86.14	21-80
	Diazoethane, 1,1,1-trifluoro	110.04	81-1000
	Dimethyl amine	45.08	5-20
	Dimethyl amine, hexafluoro	153.03	81-1000
	Dimethyl disulfide, hexafluoro	202.13	1001-10,000
20	Dimethylethylamine	73.14	21-80
	bis-(Dimethyl phosphino) amine	137.1	81-1000
	2,3-Dimethyl-2-norbornano	140.23	81-1000
	Dimethylamine, perfluoro	171.02	1001-10,000
	Dimethyloxonium chloride	82.53	21-80
25	1,3-Dioxolane-2-one, 4-methyl	102.09	81-1000
	Ethane	30.07	<5
	Ethane, 1,1,1,2-tetrafluoro	102.03	81-1000
	Ethane, 1,1,1-trifluoro	84.04	21-80
	Ethane, 1,1,2,2-tetrafluoro	102.03	81-1000
30	Ethane, 1,1,2-trichloro-1,2,2-trifluoro	187.38	1001-10,000
	Ethane, 1,1-dichloro	98	21-80
	Ethane, 1,1-dichloro-1,2,2,2-tetrafluoro	170.92	1001-10,000
	Ethane, 1,1-dichloro-1-fluoro	116.95	81-1000
	Ethane, 1,1-difluoro	66.05	5-20
35	Ethane, 1,2-dichloro-1,1,2,2-tetrafluoro	170.92	1001-10,000
	Ethane, 1,2-difluoro	66.05	5-20
	Ethane, 1-chloro-1,1,2,2,2-pentafluoro	154.47	81-1000
	Ethane, 1-chloro-1,1,2,2-tetrafluoro	136.48	81-1000
	Ethane, 2-chloro, 1,1-difluoro	100	21-80
40	Ethane, 2-chloro-1,1,1-trifluoro	118.49	81-1000
	Ethane, Chloro	64.51	5-20
	Ethane, chloro pentafluoro	154.47	81-1000
	Ethane, dichlorotrifluoro	152	81-1000
	Ethane, fluoro	48.06	5-20
45	Ethane, hexafluoro	138.01	81-1000
	Ethane, nitro-pentafluoro	165.02	81-1000
	Ethane, nitroso-pentafluoro	149.02	81-1000
	Ethane, perfluoro	138.01	81-1000
	Ethyl amine, perfluoro	171.02	1001-100,000
50	Ethyl ether	74.12	21-80
	Ethyl methyl ether	60.1	5-20
	Ethyl vinyl ether	72.11	21-80
	Ethylene	28.05	<5
	Ethylene, 1,1-dichloro	96.94	21-80
55	Ethylene, 1,1-dichloro-2-fluoro	114.93	81-1000
	Ethylene, 1,2-dichloro-1,2-difluoro	132.92	81-1000
	Ethylene, 1,2-difluoro	64	5-20

	Ethylene, 1-chloro-1,2,2-trifluoro	116.47	81-1000
	Ethylene, chloro trifluoro	116.47	81-1000
	Ethylene, dichloro difluoro	132.92	81-1000
	Ethylene, tetrafluoro	100.02	21-80
5	Fulvene	78.11	21-80
	Helium	4	<5
	1,5-Heptadiyne	92.14	21-80
	Hydrogen (H ₂)	2.02	<5
	Isobutane	58.12	5-20
10	Isobutane, 1,2-epoxy-3-chloro	106.55	81-1000
	Isobutylene	56.11	5-20
	Isoprene	68.12	5-20
	Krypton	83.8	21-80
	Methane	16.04	<5
15	Methane sulfonyl chloride, trifluoro	168.52	81-1000
	Methane sulfonyl fluoride, trifluoro	152.06	81-1000
	Methane, (pentafluorothio)trifluoro	196.06	1001-10,000
	Methane, bromo difluoro nitroso	159.92	81-1000
	Methane, bromo fluoro	112.93	81-1000
20	Methane, bromo-chloro-fluoro	147.37	81-1000
	Methane, bromo-trifluoro	148.91	81-1000
	Methane, chloro difluoro nitro	131.47	81-1000
	Methane, chloro dinitro	140.48	81-1000
	Methane, chloro fluoro	68.48	5-20
25	Methane, chloro trifluoro	104.46	81-1000
	Methane, chloro-difluoro	86.47	21-80
	Methane, dibromo difluoro	209.82	1001-10,000
	Methane, dichloro difluoro	120.91	81-1000
	Methane, dichloro-fluoro	102.92	81-1000
30	Methane, difluoro	52.02	5-20
	Methane, difluoro-iodo	177.92	1001-10,000
	Methane, disilano	76.25	21-80
	Methane, fluoro	34.03	<5
	Methane, iodo-	141.94	81-1000
35	Methane, iodo-trifluoro	195.91	1001-10,000
	Methane, nitro-trifluoro	115.01	81-1000
	Methane, nitroso-trifluoro	99.01	21-80
	Methane, tetrafluoro	88	21-80
	Methane, trichlorofluoro	137.37	81-1000
40	Methane, trifluoro	70.01	5-20
	Methanesulfonylchloride, trifluoro	136.52	81-1000
	2-Methyl butane	72.15	21-80
	Methyl ether	46.07	5-20
	Methyl isopropyl ether	74.12	21-80
45	Methyl nitrite	61.04	5-20
	Methyl sulfide	62.13	5-20
	Methyl vinyl ether	58.08	5-20
	Neon	20.18	<5
	Neopentane	72.15	21-80
50	Nitrogen (N ₂)	28.01	<5
	Nitrous oxide	44.01	5-20
	1,2,3-Nonadecane tricarboxylic acid, 2-hydroxytrimethylester	500.72	> 100,000
	1-Nonene-3-yne	122.21	81-1000
55	Oxygen (O ₂)	32	<5
	1,4-Pentadiene	68.12	5-20
	n-Pentane	72.15	21-80

	Pentane, perfluoro	288.04	> 100,000
	2-Pentanone, 4-amino-4-methyl	115.18	81-1000
	1-Pentene	70.13	5-20
	2-Pentene {cis}	70.13	5-20
5	2-Pentene {trans}	70.13	5-20
	1-Pentene, 3-bromo	149.03	81-1000
	1-Pentene, perfluoro	250.04	10,001-100,000
	Phthalic acid, tetrachloro	303.91	> 100,000
	Piperidine, 2,3,6-trimethyl	127.23	81-1000
10	Propane	44.1	5-20
	Propane, 1,1,1,2,2,3-hexafluoro	152.04	81-1000
	Propane, 1,2-epoxy	58.08	5-20
	Propane, 2,2-difluoro	80.08	21-80
	Propane, 2-amino	59.11	5-20
15	Propane, 2-chloro	78.54	21-80
	Propane, heptafluoro-1 -nitro	215.03	1001-10,000
	Propane, heptafluoro-1 -nitroso	199.03	1001-10,000
	Propane, perfluoro	188.02	1001-10,000
	Propene	42.08	5-20
20	Propyl, 1,1,1,2,3,3-hexafluoro-2,3-dichloro	221	10,001-100,000
	Propylene, 1 -chloro	76.53	21-80
	Propylene, 1-chloro-{trans}	76.53	5-20
	Propylene, 2-chloro	76.53	5-20
	Propylene, 3-fluoro	60.07	5-20
25	Propylene, perfluoro	150.02	81-1000
	Propyne	40.06	5-20
	Propyne, 3,3,3-trifluoro	94.04	21-80
	Styrene, 3-fluoro	122.14	81-1000
	Sulfur hexafluoride	146.05	81-1000
30	Sulfur (di), decafluoro(S ₂ F ₁₀)	298	> 100,000
	Toluene, 2,4-diamino	122.17	81-1000
	Trifluoroacetoneitrile	95.02	21-80
	Trifluoromethyl peroxide	170.01	81-1000
	Trifluoromethyl sulfide	170.07	81-1000
35	Tungsten hexafluoride	298	> 100,000
	Vinyl acetylene	52.08	5-20
	Vinyl ether	70	5-20
	Xenon	131.29	81-1000

Example 5. The relationship between a
 calculated Q-value for a given gas and the
 persistence of microbubbles of that gas was studied
 to determine what Q-value would be a lower limit for
 utility as an ultrasound contrast agent. For these
 experiments, a 190 x 100 mm PyrexTM (No. 3140)
 evaporation dish was filled with approximately 2000
 mL of water at 37°C. Five mL of a 20% sorbitol
 solution was taken up in a 10 mL syringe connected to
 a three way stopcock. A 10 mL syringe, containing 2
 cubic centimeters of the subject gas (or low boiling

liquid, as pertinent) was attached to the syringe containing the sorbitol solution. The sorbitol and gas or liquid are rapidly mixed 25-times to create a suspension of microbubbles or dispersed liquid and then rapidly added to the water. The microbubbles of this method are generally about 100 microns in size, and if composed of air would have a calculated persistence of 31 sec (0.5 min). Ultrasound scans before, during and after the addition were made with a Hewlett-Packard Model Sonos 500 ultrasound scanner operating at 5 MHz. The time during which the microbubbles could be observed was recorded. The results are contained in Table V below. The experimental Q-values were obtained by dividing the measured persistence of a given gas by the measured persistence for air.

TABLE V

RELATIONSHIP BETWEEN Q-VALUE FOR A GAS
AND THE PERSISTENCE OF MICROBUBBLES

GAS	Q-VALUE (Calculated)	PERSISTENCE (Experimental Q-Value)
Diethyl ether	0.1	0.1 min (0.2)
Air	1	0.6 min (1.0)
Butane	5	1.5 min (2.6)
Helium	5	2.0 min (3.5)
Propane	30	3.2 min (6.0)
Pentane	58	20.6 min (36)
Dodecafluoropentane	207,437	>5760 min (>10,105)

These experiments indicate an excellent agreement between the calculated Q-value and the experimentally determined values. Based on these data, gases with Q-values calculated to be greater than five should be potentially useful as contrast agents for ultrasound imaging.

Example 6. The relationship of the state of matter of a given chemical entity with a high Q-

coefficient and its utility as an ultrasound contrast agent was tested by comparing the efficiency of perfluoropentane and perfluorohexane to act as an ultrasound contrast agent. Perfluoropentane (dodecafluoropentane) has a calculated Q-coefficient of 207,437 and a boiling point under standard pressure conditions of 29.5 degrees Centigrade. Perfluorohexane (PCR, INC., Gainesville, FL) has a calculated Q-coefficient of 1,659,496 and a boiling point under standard pressure conditions of 59-60 degrees C. Therefore, at 37 degrees C, the body temperature of man, perfluoropentane is a gas while perfluorohexane is a liquid.

Aqueous dispersions of perfluoropentane and perfluorohexane (2% w/v) were formed at 4 degrees C by vigorous homogenization. A plastic beaker, containing approximately 1000 mL of water at 37 degrees C, was prepared to simulate human blood and was ultrasonically-scanned, as indicated in Example 5 above, before and after the addition of samples of each of the above dispersions.

Less than 1.0 mL of the perfluoropentane dispersion, when mixed with the simulated blood, produced an extremely bright ultrasound signal which persisted for at least 30 minutes. A 1:10,000 dilution was still detectable.

In contrast, a 1.0 mL sample of the perfluorohexane dispersion was undetectable by ultrasound scanning under the same conditions, as was even a 10 mL sample (1:100 dilution).

The conclusion to be drawn is that both a high Q-coefficient and a gaseous state at the body temperature of the organism being scanned is necessary for a substance to be effective as an

I claim:

1. Contrast media for ultrasound image-enhancement comprising microbubbles of a biocompatible gas having a Q coefficient greater than 5 where

$$Q = 4.0 \times 10^{-7} \times \rho / C_s D$$

and ρ is the density of the gas (Kgm^{-3}), C_s is the water solubility of the gas (M) and D is the diffusivity of the gas in solution ($\text{cm}^3\text{sec}^{-1}$).

2. Contrast media of claim 1 comprising a suspension of gas bubbles smaller than 8 microns in a biocompatible aqueous liquid vehicle.

3. Contrast media of claim 1 wherein the gas is sulfur hexafluoride.

4. Contrast media of claim 1 wherein the gas is hexafluoropropylene.

5. Contrast media of claim 1 wherein the gas is octafluoropropane.

6. Contrast media of claim 1 wherein the gas is hexafluoroethane.

7. Contrast media of claim 1 wherein the gas is octafluoro-2-butene.

8. Contrast media of claim 1 wherein the gas is hexafluoro-2-butyne.

9. Contrast media of claim 1 wherein the gas is hexafluorobuta-1,3-diene.

10. Contrast media of claim 1 wherein the gas is octafluorocyclobutane.

11. Contrast media of claim 1 wherein the gas is decafluorobutane.

5 12. Contrast media of claim 1 where the gas is dodecafluoropentane.

13. A method for selecting a gas for use as an ultrasound image-enhancement agent comprising the steps of

10 determining the solubility, C_s , of the gas in a solution;

determining the density, ρ , of the gas;

determining the diffusivity, D , of the gas in the solution;

15 calculating a Q coefficient where

$$Q = 4.0 \times 10^{-7} \times \rho / C_s D$$

and selecting a gas having a Q coefficient of greater than 5.

20 14. The method of claim 1 wherein the diffusivity, D , is determined from the molar volume, V_m , of a gas by the formula

$$D = 13.26 \times 10^{-5} \cdot \eta^{-1.14} \cdot V_m^{-.589}$$

where η is the solution viscosity (cP).

ABSTRACT

Disclosed herein are agents for enhancing the contrast in an ultrasound image. These agents are extremely small bubbles, or "microbubbles," comprised of specially selected gases. The microbubbles described herein exhibit long life spans in solution and may be produced at a size small enough to traverse the lungs, thus enabling improved ultrasound imaging of the cardiovascular system and other vital organs. Also disclosed herein is a method for selecting gases from which contrast agents may be produced. The method is based on calculations using inherent physical properties of gases and describes a means to associate the properties of a gas with the time for dissolution of microbubbles comprised of the gas.

**COMBINED DECLARATION
AND POWER OF ATTORNEY**

FOR PATENT APPLICATION

Atty

cket No. SNUS-00160

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

GASES ULTRASOUND CONTRAST MEDIA AND METHOD FOR SELECTING GASES FOR USE AS ULTRASOUND CONTRAST MEDIA

the specification of which was filed as PCT International Application No. PCT/US92/_____ on September __, 1992.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)

Priority Claimed
Yes No

Number Country Day/Month/Year Filed

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose all information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Application Ser. No.	Filing Date	Status: Patented, Pending, Abandoned
07/893,657	June 5, 1992	Pending
07/761,311	September 17, 1991	Pending

I HEREBY APPOINT THE FOLLOWING AS MY ATTORNEYS WITH FULL POWER OF SUBSTITUTION TO PROSECUTE THIS APPLICATION AND TRANSACT ALL BUSINESS IN THE PATENT OFFICE CONNECTED THEREWITH:

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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